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# Introduction of Water-Solubilizing Groups at the Lower Rim of Tolylpyridine-Bridged Cavitands

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In this paper the preparation of water-soluble methylene-bridged cavitands presenting either positively or negatively charged groups at the lower rim following three different synthetic routes is reported. Moreover, four anionic sulphate functions have been successfully inserted on a tolylpyridine-bridged cavitand in order to carry out the self-assembly of coordination cages in water.

*Keywords*: Resorcinarenes; Water-soluble cavitands; Coordination cages; deep-cavity cavitands

#### INTRODUCTION

Water-soluble cavitands are interesting molecular receptors thanks to their versatile complexation abilities [1,2]. In parallel, cavitands have been extensively studied as tetradentate ligands for the self-assembly of coordination cages in organic solvents [3,4]. Water solubility of the coordination cages is an essential element to exploit the peculiar properties of their interiors. An outstanding example of water-soluble cavitand-based capsules, selfassembled through metal coordination, has been reported by the Harrison group [5,6]. Capsule formation and guest encapsulation are triggered by pH changes. The iminodiacetate ligands are attached to the cavitand upper rim and at the same time are responsible for the water solubility of the otherwise hydrophobic cavitand. Addition of cobalt (II) or iron (II) ions at pH > 5 results in the formation of the water soluble cages with the hydrophilic groups and metal ions positioned in the equatorial part of the cage. An alternative strategy worth exploring, is to assemble cavitand-based coordination cages moving the water-solubilizing groups from the equatorial

zone to the polar zone, as sketched in Fig. 1. This requires incorporating the water-solubilizing groups at the lower rim of the cavitand tetradentate ligand, leaving the upper apical and bridge positions free for ligand introduction. In our case the ligands responsible for metal coordination are tolylpyridine bridging units appended to the upper rim [3,7] as separate entities from the hydrophilic groups. By adding two equivalents of a suitable metal precursor to a water solution of these cavitands it would be possible to assemble a water-soluble nanosized cage directly in aqueous solution (Fig. 1).

Water soluble cavitands bearing charged groups at the lower rim have been described in the literature. The water-solubilizing groups chosen were ammonium salts, [8] phosphates [9], and carboxylates [10]. All these cavitands have been synthesized from a common precursor, the hydroxylfooted resorcinarene **9** [11].

In this paper we report the preparation of watersoluble methylene-bridged cavitands presenting either positively ( $NH_3^+$ , **4b**, **7b**) or negatively charged groups ( $OSO_3^-$ , **14**) at the lower rim, the synthesis of the tetradentate cavitand ligand **15** having four sulphate feet and attempts to form the corresponding coordination cages.

In order to work out the best synthetic conditions for the introduction of the charged groups at the lower rim, we synthesized the methylene-bridged cavitands **1**, **5** and **13** as model compounds by applying the method reported by Kaifer for the bridging reaction [12]. For the introduction of the positively charged ammonium groups two different strategies were attempted, as reported in Schemes 1 and 2. Both gave the desired compounds **4a** and **7a** 

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FIGURE 1 Sketch of cage self-assembly.

which turned out to be completely water soluble under acidic conditions. In the procedure of Scheme 1 the lower rim hydroxyls of 1 were converted to NH<sub>2</sub> groups with a 33% overall yield. Specifically, the azide derivative **3** was prepared following a synthetic procedure reported by Rebek et al. [8] by mesylation of **1** with MsCl (Et<sub>3</sub>N, CH<sub>2</sub>Cl<sub>2</sub>, 91% yield) and further treatment with NaN<sub>3</sub> in DMF at 70°C (52% yield). The one-pot reduction of **3** using PPh<sub>3</sub> in THF/H<sub>2</sub>O at 70°C resulted in cavitand **4a** which was then transformed in the tetraammonium salt **4b** in acidic water (10% aq. HCl) (Scheme 1). The resulting cavitand is water-soluble.

Alternatively, **5**, having four bromo substituents in the apical positions, was submitted to Mitsunobu reaction with phthalimide, diethyl azodicarboxylate (DEAD), and PPh<sub>3</sub> in THF (78% yield) followed by the Gabriel reaction with hydrazine in ethanol (92% yield) resulting in tetraamine functionalized cavitand **7a**, whose ammonium salt is completely soluble in H<sub>2</sub>O (Scheme 2). This second procedure gave an overall yield of 72% from **5**, more than doubled if compared to the overall yield of the first synthetic pathway (33 %).

For this reason the Mitsunobu–Gabriel reaction was selected as the chosen method for the synthesis of the tolylpyridine-bridged cavitand **12** with four amine at the lower rim (Scheme 3). The synthesis of the hydroxyl-footed tetradentate cavitand **10** was performed by bridging the corresponding resorcinarene **9** with dibromotolylpyridine in dry DMA at 80°C (30% yield). Then, as in the previous case, the hydroxyls were converted into phthalimide groups giving cavitand **11** with a 31% yield. The subsequent reaction with hydrazine in ethanol resulted in the desired tetraamine cavitand **12** (85% yield). Unaccountably, the following step of salification in 10% aqueous HCl to give the corresponding water soluble

tetrammonium salt failed. The HCl added caused the breaking of the tolylpyridine bridges as demonstrated by the recovery of the corresponding tolylpyridine aldehyde and tetraamine resorcinarene in the aqueous solution. In fact ESI-MS spectrometry showed the presence of two prominent peaks at 184 and 717 m/z due to these two compounds. Water-solubility through acidification of the amine groups at the lower rim cannot be applied to the case of our tetradentate cavitand ligand, due to the reactivity of the benzal-type bridges. For this reason the introduction of positively charged ammonium groups at the lower rim is not a valuable strategy for the construction of water-soluble pyridyl tetradentate ligands.

The second approach to the introduction of hydrophilic groups at the lower rim of tetradentate cavitands was through the functionalization with sulphate anions. Once again the first synthetic effort has been focused on the synthesis of the methylenebridged cavitand as model compound. At first the synthesis of tetrasulphonate water soluble resorcinarenes reported by Aoyama [13] was attempted, but the following bridging reaction failed due to the presence of the sulphonate groups.

For this reason we moved to another strategy: the introduction of the hydrophilic sulphate groups on a preformed cavitand. In a recent article, Linhardt [14] reported an interesting way to functionalize sugars with sulphate groups in order to increase their solubility in water. This method consists in the sulphonation of the OH groups using NMe<sub>3</sub>·SO<sub>3</sub> in DMF at 50°C. Cavitand **13** was therefore combined with sulphur trioxide trimethylamine complex giving the water soluble tetrasulphate cavitand **14** with a 65% yield (Scheme 4).

The success of this approach on the model compound encouraged us to follow the same protocol on tolylpyridine-bridged cavitand **10** 



SCHEME 1 Reagents and conditions: a) MsCl, CH<sub>2</sub>Cl<sub>2</sub>/Et<sub>3</sub>N, 0°C, 3 h, 91%; b) NaN<sub>3</sub>, DMF, 70°C, 24 h, 52%; c) PPh<sub>3</sub>, THF/H<sub>2</sub>O, 70°C, 2 h, 88%; d) 10% aq. HCl.

which was reacted with NMe<sub>3</sub>·SO<sub>3</sub> as previously described (Scheme 5).

The insertion of the four sodium sulphate groups was successful as confirmed by ESI-MS and <sup>1</sup>H NMR analyses. Unfortunately cavitand **15** is not soluble in water, since the presence of four tolylpyridine groups as bridging units make the hydrophobic part of the molecule predominant despite, the presence of four charged sulphate salts. Cavitand **15** is soluble in water/alcohol and water/DMSO mixtures.

As already proven by Fujita, water solubility is not strictly necessary for the organic ligands if the metal precursor is sufficiently hydrophilic to make the resulting nanosize cage water-soluble [15]. Cavitand **15** was therefore tested for the self-assembly of the corresponding coordination cage in the presence of Pd(en)(NO<sub>3</sub>)<sub>2</sub>, exploiting the eight nitrate counterions to promote the water solubility of the cage. The self-assembly led to the formation of insoluble precipitates. The outcome is the same either starting from a homogeneous solution (DMSO) or from a cavitand **15** suspension in water. Different working temperatures do not change the situation. At this stage of the work we can only speculate about the failure of cage self-assembly. Our hypothesis is the following: the  $R-O-SO_3^-$  groups displace the counterions of the metal precursor interfering with the self-assembly process [16].

#### **CONCLUSIONS**

In summary, three different synthetic routes to obtain water soluble methylene-bridged cavitands have been reported.



SCHEME 2 Reagents and conditions: a) Phthalimide, PPh<sub>3</sub>, DEAD, THF, reflux, 5 days, 78%; b) NH<sub>2</sub>NH<sub>2</sub>, EtOH, reflux, 3 h, 92%; c) 10% aq. HCl.

In the case of the tolylpyridine-bridged cavitands, acidification with 10% aqueous HCl of tetraamine-footed cavitand **12** led to the breaking of the bridging ligand groups, thus precluding the following step of cage self-assembly. On the other hand the insertion of four anionic sulphate functions on the same tetradentate cavitand turned out to be synthetically viable, but not sufficient to promote the solubility of the cavitand in water. Water-solubility in these type of cavitands is heavily dependent on the size of the hydrophobic bridging units. Cage self-assembly has shown to be inhibited by the presence of anionic groups at the lower rim.

#### **EXPERIMENTAL**

#### **General Methods**

All commercial reagents were ACS grade and used as received. All solvents were dried over 3 Å and 4 Åmolecular sieves. <sup>1</sup>H NMR spectra were recorded on Bruker AC300 (300 MHz), Avance (300 MHz) and AMX400 (400 MHz) spectrometers and all chemical shifts ( $\delta$ ) were reported in parts per million (ppm) in relation to the proton resonances which resulted from incomplete deuteration of the NMR solvents. Mass spectra of the organic compounds were measured with a Finnigan MAT SSQ710 spectrometer, using the CI (chemical ionization) technique. Electrospray ionization mass spectrometry (ESI-MS) experiments were performed on a Waters ZMD spectrometer equipped with an electrospray interface. Matrix-assisted laser desorption ionization time-of-flight (MALDI-TOF) mass spectra were obtained on a PerSpective Biosystems Voyager DERP spectrometer equipped with delayed extraction. Column chromatography was performed by using silica gel 60 (Merck 70–230 mesh).

Cavitands **1** [9], **5** and **13** [17], and 4,4'-(a,a'-dibromo)tolylpyridine **8** [7], were prepared according to literature procedures.

#### Cavitand 2

Methanesulfonylchloride  $(25 \,\mu\text{L}, 0.320 \,\text{mmol})$  was added drop by drop to a stirred solution of **1** (56 mg, 0.067 mmol) in CH<sub>2</sub>Cl<sub>2</sub> (10 mL) and Et<sub>3</sub>N (0.12 mL, 0.870 mmol) at 0°C. After 3 h of stirring at 0°C, the solution was poured into 10% aq. HCl. The aqueous layer was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed with aqueous NaHCO<sub>3</sub> solution and concentrated to obtain the corresponding cavitand **2** (69 mg, 91%).



SCHEME 3 Reagents and conditions: a) DMA,  $K_2CO_3$ , 80°C, 48 h, 30%; b) Phthalimide, PPh<sub>3</sub>, DEAD, THF, reflux, 5 days, 31%; c)  $NH_2NH_2$ , EtOH, reflux, 3 h, 85%.



SCHEME 4 Reagents and conditions: a) NMe<sub>3</sub>·SO<sub>3</sub>, DMF, 50°C, 24 h, 65%.



SCHEME 5 Reagents and conditions: a) NMe<sub>3</sub>·SO<sub>3</sub>, DMF, 50°C, 24 h, 31%.

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz):  $\delta = 7.05$  (s, 4H, ArH<sub>down</sub>); 5.86 (d, 4H, OCH<sub>out</sub>O, <sup>2</sup>J = 7.0 Hz); 4.79 (t, 4H, ArCH, <sup>3</sup>J = 8.0 Hz); 4.36 (t, 8H, CH<sub>2</sub>OMs, <sup>3</sup>J = 6.3 Hz); 4.26 (d, 4H, OCH<sub>in</sub>O, <sup>2</sup>J = 7.0 Hz); 2.95 (s, 12H, CH<sub>3</sub>S); 2.39 (m, 8H, ArCHCH<sub>2</sub>); 1.96 (s, 12H, ArCH<sub>3</sub>); 1.79 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

MALDI TOF-MS (m/z): 1161  $[M + Na]^+$ .

#### Cavitand 3

To a solution of **2** (69 mg, 0.060 mmol) in DMF (2 mL) was added NaN<sub>3</sub> (23 mg, 0.350 mmol) and the mixture was stirred for 24 h at 70°C. After, the reaction mixture was poured into aqueous Na<sub>2</sub>CO<sub>3</sub>, and the formed precipitate was filtered, washed with water and dried to give **3** (29 mg, 52%).

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz):  $\delta = 6.96$  (s, 4H, ArH<sub>down</sub>); 5.88 (d, 4H, OCH<sub>out</sub>O, <sup>2</sup>J = 7.4 Hz); 4.79 (br t, 4H, ArCH<sub>i</sub>); 4.24 (d, 4H, OCH<sub>in</sub>O, <sup>2</sup>J = 7.4 Hz); 3.42 (br t, 8H, CH<sub>2</sub>N<sub>3</sub>); 2.32 (m, 8H, ArCHCH<sub>2</sub>); 1.96 (s, 12H, ArCH<sub>3</sub>); 1.66 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

CI-MS (m/z): 925  $[M]^+$ , 898  $[M-N_2]^+$ .

#### Cavitand 4b

PPh<sub>3</sub> (83 mg, 0.317 mmol) was added to a solution of **3** (29 mg, 0.032 mmol) in THF (2 mL) and water (40  $\mu$ L, 2.220 mmol). The mixture was stirred for 2 h at 70°C and then poured into aqueous Na<sub>2</sub>CO<sub>3</sub>. The aqueous layer was extracted with EtOAc and the organic layer was concentrated in *vacuo* to give **4a** (23 mg, 88%). The free amine **4a** was converted into the corresponding tetraammonium salt **4b** by using 10% aq. HCl.

<sup>1</sup>H NMR (D<sub>2</sub>O, 300 MHz):  $\delta = 7.38$  (s, 4H, ArH<sub>down</sub>); 5.87 (d, 4H, OCH<sub>out</sub>O, <sup>2</sup>J = 7.4 Hz); 4.61 (t, 4H, ArCH, <sup>3</sup>J = 8.1 Hz); 4.09 (d, 4H, OCH<sub>in</sub>O,

 ${}^{2}J = 7.4 \text{ Hz}$ ; 3.02 (t, 8H, CH<sub>2</sub>NH<sub>2</sub>,  ${}^{3}J = 7.5 \text{ Hz}$ ); 2.47 (m, 8H, ArCHCH<sub>2</sub>); 1.82 (s, 12H, ArCH<sub>3</sub>); 1.58 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

4a: ESI-MS (m/z): 823  $[M + H]^+$ , 412  $[M + 2H]^{2+}$ .

**4b**: C<sub>48</sub>H<sub>64</sub>Cl<sub>4</sub>N<sub>4</sub>O<sub>8</sub> (966.86): calcd. C 59.63, H 6.67, N 5.79; found C 59.23, H 6.74, N 5.66.

#### Cavitand 6

Phthalimide (120 mg, 0.815 mmol), PPh<sub>3</sub> (213 mg, 0.812 mmol) and DEAD (257 mg, 1.475 mmol) were added to a solution of **5** (200 mg, 0.184 mmol) in dry THF (60 mL). The mixture was refluxed for 5 days. The reaction was quenched in brine (30 mL) and the aqueous solution extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was concentrated in *vacuo*, and the residue was recrystalized from CH<sub>2</sub>Cl<sub>2</sub> and EtOH to give **6** (230 mg, 78%).

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz):  $\delta = 7.73$  (m, 8H, *H<sub>Fl</sub>*); 7.59 (m, 8H, *H<sub>Fl</sub>*); 7.18 (s, 4H, Ar*H<sub>down</sub>*); 5.92 (d, 4H, OC*H<sub>out</sub>*O, <sup>2</sup>*J* = 7.4 Hz); 4.89 (t, 4H, ArC*H*, <sup>3</sup>*J* = 8.0 Hz); 4.36 (d, 4H, OC*H<sub>in</sub>*O, <sup>2</sup>*J* = 7.4 Hz); 3.91 (t, 8H, CH<sub>2</sub>N, <sup>3</sup>*J* = 7.2 Hz); 2.42 (m, 8H, ArCHCH<sub>2</sub>); 1.79 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

CI-MS (m/z): 1602  $[M + H]^+$ .

#### Cavitand 7a

To the solution of **6** (200 mg, 0.125 mmol) in EtOH (10 mL) hydrazine hydrate was added (7.3 mL, 0.149 mol) and the mixture was refluxed under nitrogen atmosphere for 3 h. The EtOH was evaporated and the residue taken up in 20 mL of 2N aqueous KOH solution. The basic solution was extracted with  $CH_2Cl_2$ . The organic layer was

washed with brine, dried, and concentrated in *vacuo* to give **7a** (125 mg, 92%).

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz):  $\delta = 7.16$  (s, 4H, ArH<sub>down</sub>); 5.95 (d, 4H, OCH<sub>out</sub>O, <sup>2</sup>J = 7.2 Hz); 4.84 (t, 4H, ArCH, <sup>3</sup>J = 7.8 Hz); 4.38 (d, 4H, OCH<sub>in</sub>O, <sup>2</sup>J = 7.2 Hz); 2.80 (br t, 8H, CH<sub>2</sub>NH<sub>2</sub>); 2.32 (br t, 8H, ArCHCH<sub>2</sub>); 1.46 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

CI-MS (m/z): 1082  $[M + H]^+$ .

**7b**: C<sub>44</sub>H<sub>52</sub>Br<sub>4</sub>Cl<sub>4</sub>N<sub>4</sub>O<sub>8</sub> (1226.33): calcd. C 43.09, H 4.27, N 4.57; found C 42.96, H 4.42, N 4.46.

#### Cavitand 10

1.580 g (4.83 mmol) of 4,4'-(a,a'-dibromo)tolylpyridine 8 and 1.150 g (8.28 mmol) of K<sub>2</sub>CO<sub>3</sub> were added under nitrogen to a solution of 0.500 g (0.69 mmol) of resorcinarene 9 in dry DMA (25 mL). The mixture was stirred in a sealed tube at 80°C for 48 h. The reaction was quenched by water addition (10 mL) and the formed precipitate was filtered. The black crude obtained was purified by column chromatography (SiO<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>/Ethanol 9:1) to give compound **10** as a yellow solid (0.302 g, 30%).

<sup>1</sup>H NMR ([D<sub>6</sub>]DMSO, 400 MHz):  $\delta = 8.65$  (d, 8H, XX' part of system AA'XX',  $H_{oPy}$ , <sup>3</sup>J = 4.8 Hz); 7.90 (d, 8H, BB' part of system AA'BB', <sup>3</sup>J = 8.0 Hz); 7.86 (d, 8H, AA' part of system AA'BB', <sup>3</sup>J = 8.0 Hz); 7.82 (s, 4H, ArH<sub>down</sub>); 7.70 (d, 8H, AA' part of system AA'XX',  $H_{mPy}$ , <sup>3</sup>J = 4.8 Hz); 6.96 (s, 4H, ArH<sub>up</sub>); 5.58 (s, 4H, OCHO); 4.82 (t, 4H, ArCH, <sup>3</sup>J = 8.0 Hz); 4.51 (t, 4H, CH<sub>2</sub>OH, <sup>3</sup>J = 4.8 Hz); 3.58 (m, 8H, CH<sub>2</sub>OH); 2.55 (m, 8H, ArCHCH<sub>2</sub>); 1.54 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>). CI-MS (m/z): 1381 [M + H]<sup>+</sup>.

#### Cavitand 11

Phthalimide (38 mg, 0.261 mmol), PPh<sub>3</sub> (68 mg, 0.261 mmol) and DEAD (83 mg, 0.475 mmol) were added to a solution of **10** (82 mg, 0.059 mmol) in dry THF (20 mL). On the second and sixth day, an additional portion of phthalimide (38 mg), PPh<sub>3</sub> (68 mg) and DEAD (83 mg) was added to the mixture. The reaction was refluxed for 8 days, quenched in brine (30 mL) and the aqueous solution extracted with diethyl ether. The organic layer was concentrated in *vacuo*, and the crude product obtained was purified by column chromatography (SiO<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>/EtOAc 7:3, CH<sub>2</sub>Cl<sub>2</sub>/EtOH 8:2) to give compound **11** (35 mg, 31%).

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 600 MHz):  $\delta$  = 8.68 (bd, 8H, XX' part of system AA'XX', *H*<sub>oPy</sub>); 7.82 (bd, 8H, BB' part of system AA'BB'); 7.73 (bd, 8H, *H*<sub>Pht</sub>); 7.70 (bd, 8H, AA' part of system AA'BB'); 7.64 (bd, 8H, *H*<sub>Pht</sub>); 7.59 (bd, 8H, AA' part of system AA'XX', *H*<sub>mPy</sub>); 7.46 (s, 4H, Ar*H*<sub>down</sub>); 6.72 (s, 4H, Ar*H*<sub>up</sub>); 5.55 (s, 4H, OCHO); 5.02 (t, 4H, ArCH, <sup>3</sup>*J* = 7.8 Hz); 4.00 (t, 8H, CH<sub>2</sub>N, <sup>3</sup>*J* = 7.2 Hz); 2.61 (m, 8H, ArCHCH<sub>2</sub>); 1.90 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

CI-MS (m/z): 1898 [M]<sup>+</sup>, 1753 [M-Pht]<sup>+</sup>, 1607 [M-2Pht]<sup>+</sup>.

#### Cavitand 12

To the solution of **11** (35 mg, 0.018 mmol) in EtOH (5 mL) hydrazine hydrate (14 mL, 0.221 mol) was added and the mixture was refluxed under nitrogen atmosphere for 3 h. The EtOH was evaporated and the residue taken up in 10 mL of 2N aqueous KOH solution. The basic solution was extracted with CH<sub>2</sub>Cl<sub>2</sub>. The organic layer was washed with brine, dried, and concentrated in *vacuo* to give **12** (22 mg, 85%).

<sup>1</sup>H NMR (CDCl<sub>3</sub>, 300 MHz):  $\delta = 8.67$  (bd, 8H, XX' part of system AA'XX', *H*<sub>oPy</sub>); 7.81 (d, 8H, BB' part of system AA'BB', <sup>3</sup>J = 8.3 Hz); 7.71 (d, 8H, AA' part of system AA'BB', <sup>3</sup>J = 8.3 Hz); 7.50 (bd, 8H, AA' part of system AA'BB', <sup>3</sup>J = 8.3 Hz); 7.33 (s, 4H, AA' part of system AA'XX', *H*<sub>mPy</sub>); 7.33 (s, 4H, ArH<sub>down</sub>); 6.74 (s, 4H, ArH<sub>up</sub>); 5.57 (s, 4H, OCHO); 4.98 (bt, 4H, ArCH); 3.71 (m, 8H, CH<sub>2</sub>NH<sub>2</sub>, <sup>3</sup>J = 7.0 Hz); 2.89 (m, 8H, ArCHCH<sub>2</sub>); 1.61 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

MALDI TOF-MS (m/z): 1377  $[M]^+$ .

C<sub>88</sub>H<sub>80</sub>N<sub>8</sub>O<sub>8</sub> (1377.63): calcd. C 76.72, H 5.85, N 8.13; found C 76.68, H 5.87, N 8.09.

#### Cavitand 14

NMe<sub>3</sub>·SO<sub>3</sub> complex (68.5 mg, 0.49 mmol) was added to a solution cavitand **13** (63 mg, 0.08 mmol) in dry DMF (10 mL). The mixture was stirred under nitrogen atmosphere at 50°C for 12 h. An additional portion of NMe<sub>3</sub>·SO<sub>3</sub> complex (68.5 mg) was added, and the suspension was kept at 50°C for another 12 h. The reaction was quenched by the addition of water (10 mL) and Na<sub>2</sub>CO<sub>3</sub> was added until the pH reached 7.After concentration, a mixture of methanol/water was added to precipitate the unreacted complex and the salts. After filtration, a mixture of acetone/water was added to the solution to precipitate cavitand **14** as light pink solid (62 mg, 65%).

<sup>1</sup>H NMR (D<sub>2</sub>O, 400 MHz): δ: 1.78 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>); 2.51 (q, 8H, ArCHCH<sub>2</sub>); 4.16 (t, 8H, CH<sub>2</sub>CH<sub>2</sub>OSO<sub>3</sub>Na, <sup>2</sup>*J* = 6.4 Hz); 4.36 (d, 4H, CH<sub>in</sub>, <sup>2</sup>*J* = 7.8 Hz); 5.8 (d, 4H, CH<sub>out</sub>, <sup>2</sup>*J* = 7.5 Hz); 6.69 (s, 4H, ArH<sub>up</sub>); 7.49 (s, 4H, ArH<sub>down</sub>).

ESI-MS (m/z): 1154 [M-Na]<sup>-</sup>, 565 [M-2Na]<sup>2-</sup>, 369 [M-3Na]<sup>3-</sup>, 271 [M-4Na]<sup>4-</sup>.

#### Cavitand 15

 $NMe_3 \cdot SO_3$  complex (60.5 mg, 0.43 mmol) was added to a solution cavitand **10** (100 mg, 0.07 mmol) in dry DMF (10 mL). The mixture was stirred under nitrogen atmosphere at 50°C for 12 h. An additional portion of  $NMe_3 \cdot SO_3$  complex (60.5 mg) was added, and the suspension was kept at 50°C for another 12 h. The reaction was quenched by the addition of water (10 mL) and Na<sub>2</sub>CO<sub>3</sub> was added until the pH reached 7. After concentration, a mixture of methanol/water was added to precipitate the unreacted complex and the salts. After filtration, a mixture of acetone/water was added to the solution to precipitate cavitand **15** as light brown solid (40 mg, 31%).

<sup>1</sup>H NMR ([D<sub>6</sub>]DMSO, 300 MHz):  $\delta = 8.64$  (d, 8H, XX' part of system AA'XX',  $H_{oPy}$ , <sup>3</sup>J = 5.9 Hz); 7.90 (bd, 8H, BB' part of system AA'BB', <sup>3</sup>J = 8.1 Hz); 7.85 (m, 12H, AA' part of system AA'BB' + ArH<sub>down</sub>); 7.70 (d, 8H, AA' part of system AA'XX',  $H_{mPy}$ , <sup>3</sup>J = 5.9 Hz); 6.93 (s, 4H, ArH<sub>up</sub>); 5.57 (s, 4H, OCHO); 4.81 (t, 4H, ArCH, <sup>3</sup>J = 8.0 Hz); 3.87 (bm, 8H, CH<sub>2</sub>OSO<sub>3</sub>Na); 2.56 (m, 8H, ArCHCH<sub>2</sub>); 1.64 (m, 8H, CH<sub>2</sub>CH<sub>2</sub>CH<sub>2</sub>).

ESI-MS (m/z): 1767 [M-Na]<sup>-</sup>, 872 [M-2Na]<sup>2-</sup>, 573 [M-3Na]<sup>3-</sup>, 424 [M-4Na]<sup>4-</sup>.

C<sub>88</sub>H<sub>72</sub>N<sub>4</sub>Na<sub>4</sub>O<sub>24</sub>S<sub>4</sub> (1789.74): calcd. C 59.06, H 4.05, N 3.13; found C 58.97, H 4.16, N 3.01.

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74